HANDBOOK

Rapid Prep 96 Plasmid Kit

☐	**Nucleo-pore™ Rapid Prep 96 Plasmid Kit**	NP-36172	4 x 96 Preps
☐	**Nucleo-pore™ Rapid Prep 96 Plasmid Kit**	NP-36174	24 x 96 Preps

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COMPONENTS

KIT CONTENTS

Nucleo-pore™ Rapid Prep 96 Plasmid kit

Cat. No.	NP-36172	NP-36174
NO. of Preps.	4x96	24x 96
Buffer R1-96	250 ml	6 x 250 ml
Buffer R2-96	250 ml	6 x 250 ml
Buffer R3-96	250 ml	6 x 250 ml
Buffer RE-96	60 ml	6 x 60 ml
RNase A (lyophilized)	50 mg	6 x 50 mg
RC-Plate (culture plate)	4	24
RP-Plate (precipitation plate)	4	24
RGP-Foil	4	24
RSA-Foil	20	120
RF-Plates (Filter Plates)	4	24
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The kit for 24 x 96 preparations (Cat. No. NP-36174) consists of 6 x Cat. No.NP-36172

Reagents, consumables and equipments not provided with the kit

- Centrifuge with a swinging-bucket rotor attaining ≥ 2,500 × g (for harvesting of bacterial cells and plasmid precipitation). Specifications: Clearance of the buckets must be sufficient to accommodate RP-Plate (height: 44 mm).
- Vacuum manifold (for clearing of the neutralized and heat-incubated lysate) required for manual use of the Nucleo-pore Rapid Prep 96 plasmid kit.
- For use with a centrifuge with a swing-out rotor capable to accommodate the RF-Plate/RP-Plate sandwich (bucket height: 85 mm), e.g. Hermle Z 513/Z 513 K, Jouan KR4i, Heraeus Kendro Multifuge 3/3-R, Beckman Coulter Allegra 25R, Hettich Rotanta 460 series, Sigma 4-15/4K15/6-15/6K15 the whole procedure can be performed in the centrifuge. For centrifuges, capable of attaining 6,000 × g, centrifugation times can be further reduced and optimized.
- Compatible automated workstations for Nucleo-pore Rapid Prep 96 plasmid kit : Biomek 2000/FX (Beckman-Coulter); MiniPrep series (Cavro); Microlab Star (Hamilton); RoboSmart/RoboPrep (MWG); MultiPROBE II/II HT (Perkin Elmer); BioRobot 9600/3000/8000 (Qiagen); Genesis RSP/RWS Separation System series (Tecan); SciClone ALH (Zymark).

SAFETY INSTRUCTIONS

Always wear gloves and goggles and follow the Safety Information.

RNaseA

Contains lyophilized RNaseA. R&S Phrases: R 42/43; S 22-24

Buffer R2-96

Contain Sodium hydroxide (<2%). R&S Phrases: R 36/38, S 26-37, 39-45

R & S Phrases

R 36/38 Irritating to eyes and skin, R 42/43 May cause sensitization by inhalation and skin contact S 22 Do not breathe dust; S 24 Avoid contact with the skin; S 26 In case of contact with eyes, rinse immediately with plenty of water and seek medical advice; S 37/39 Wear suitable gloves and eye/face protection; S 45 In case of accident or if you feel unwell, seek medical advice immediately

INTRODUCTION

Principle and Procedure

Buffers R1-96 and Buffer R2-96 efficiently liberates plasmid DNA from E.coli host cells The resulting lysate is then neutralized by buffer R3-96. Inactivation of nucleases is done by Incubation in boiling water (optional). Precipitates are removed by filtration through RF-Plate under vacuum or in a suitable centrifuge for microtiter plates. Plasmid DNA is then precipitated by addition of isopropanol to the filtrate followed by centrifugation. After an additional washing step with 70% ethanol and drying, the DNA can be resuspended in buffer RE-96.

Specification of Nucleo-pore Rapid Prep 96 Plasmid Kit

- Nucleo-pore Rapid Prep 96 Plasmid kit is designed for the rapid manual and automated 96- well DNA preparation of high and low copy-number plasmids and large lowcopy constructs (e.g. BACs, bacterial artificial chromosomes) DNA from E. coli cultures.
- Upto 8 µg of highly pure plasmid DNA from 1.2ml of a saturated E. coli culture (per preparation) can be purified in the convenient 96-well format.
- The prepared plasmid DNA is suitable for several downstream applications including automated fluorescent DNA sequencing applications, PCR, or enzymatic manipulation.
- Time for manual parallel processing of up to 384 plasmid DNA minipreps from E. coli cultures with Nucleo-pore Rapid Prep 96 Plasmid kit is less than 90 min.

FEATURES

- Culture volume 1.1-1.3 ml
- Average yield 8 µg (plasmids); 0.2 µg (BACs);
- Vectors up to 250 kb
- Time/prep 1hr 30min with 4 x 96

Compatibility with common vacuum manifolds

The Nucleo-pore Rapid Prep 96 Plasmid kit can be used with QIAvac 96 (Qiagen) without need of any additional equipment. Howerver, it is not suitable for BioRad/Aurum, Eppendorf/Perfect VAC or Millipore/MultiScreen vacuum manifolds.

Preparation and storage of Reagents

Keep all the bottles tightly closed at all times. Buffer R2-96 can absorb carbon dioxide when exposed to air resulting in decreased pH.

If a precipitate is observed in Buffer R2-96 (as SDS may precipitate if stored below 18ºC), incubate the bottle at 30 – 40°C for some minutes and mix well until all of the precipitation is re-dissolved.

If using the Nucleo-pore Rapid Prep 96 Plasmid kit manually, establish a reliable vacuum source for the compatible vacuum manifold. The manifold may be used with a vacuum pump, house vacuum, or water aspirator. However, a vacuum of 200-400 mbar (pressure difference) is recommended. The use of the manifold with vacuum regulator is recommended.

RNaseA

Reconstitute RNaseA in Buffer R1-96 by adding 1 ml Buffer R1-96 to the RNase A vial, vortex, and pipette all of the resulting solution to the Buffer R1-96 bottle. The RNase in the mixture is stable for six months if stored at +4°C.

BACTERIAL CULTURE

Culture medium

LB (Luria-Bertani) medium
Conditions : Cultivate cells at 37°C with constant shaking at 250 rpm.
2 x YT or Terrific Broth (TB) can be used.
(in2 x YT or TB, bacteria grow faster and reach the stationary phase much earlier than in LB medium i.e. less than12 h).

Cultivation in RC-Plate

Autoclave the RC-Plates (supplied) or use a sterile block. Add 1.2-1.5 ml (do not exceed 1.5ml as it may cause spillage) of selected medium (with antibiotic, e.g. 100 µg/ml ampicillin) to each well. Inoculate each well with a single bacterial colony. Cover the RC-Plates with the RGP-Foil (supplied). Grow the culture at 37°C for 16-24 h in an incubator with vigorous shaking (200-400 rpm). The RC-Plate may be fixed to the shaker with large-size flask clamps or tape.

Note:
It is recommended to use high copy- number plasmids such as pBluescript or pGEM, pUC and E. coli strains like DH5α or XL1 Blue.

Cultivation in Tubes

Depending on the bacterial strain and copy number of the plasmid 1 to 5 ml LB medium (do not exceed5ml) or 3 ml 2 x YT or 3 ml TB medium (do not exceed 3ml) can be used. Shake vigorously for 10-14 h.
Optional:
Transfer bacterial culture from the tubes to the RC-Plates supplied with the kit. For this, transfer 1.5 ml of the culture to each well of the RC-Plate. Harvest the cultures by centrifugation. Discard supernatant. Usually 1.5 ml of culture is sufficient for DNA preparation. However, if necessary, add an additional 1.0-1.5 ml of bacterial culture to each well of the RC-Plate centrifuge again & discard the supernatant.

PROTOCOL

Manual purification of high-copy plasmid DNA

Procedure

1. Centrifuging the RC-Plates to pellet bacteria for 10 min at 1,000 × g (leave RGP-Foil on the block) . Remove the RGP-Foil and invert the block quickly to pour off the supernatant. Tap the block on a paper sheet or paper towels in order to ensure maximum removal of the remaining medium.

2. Add 300 µl Buffer R1-96 (RNaseA added) to each bacterial pellet and vortex. until no more cell clumps or pellets are visible.

3. Add 300 µl Buffer R2-96 to each well. Seal the block thoroughly with RSA-Foil. Gently press the foil all over so that it properly sticks leaving no air space between wells in order to to prevent well-to-well cross contamination. Mix by gently inverting the block 3 – 4 times. Incubate at room temperature for up to 5 min (do not exceed 5 minutes). Do not vortex as vortexing will shear the DNA, leading to contamination of the resulting plasmid DNA preparation. Remove RSA-Foil gently and dry the upper rim of the wells with a lint-free tissue paper.
   Do not extend the cell lysis to more than 5 min.

4. Add 300 µl R3-96 to each well. Seal the block thoroughly with RSA-Foil Gently press the foil all over so that it properly sticks leaving no air space between wells in order to to prevent well-to-well cross contamination. Mix by gently inverting the block 3 – 4 times.
   A white precipitate of SDS and cell debris forms.

5. (Optional Step for endA+ host strains): Seal the block with a fresh RSA-Foil and incubate it in a boiling water bath for 5 min for the heat destruction of endonucleases and denaturation of proteins. Let the block cool down on ice to at least room temperature.
   Note: This heating step can be omitted when using end A- host strains like DH5α or XL-1 Blue.

Adjust the vacuum manifold:
Insert spacers for “RP-Plates”, , into the grooves located on the short sides of the manifold with notched side up. Place a fresh RP-Plate into the manifold base, close the lid and insert the RF-Plate into the lid.
If using a centrifuge with a swing-bucket rotor capable of hosting 96-well blocks plus the RF-Plate, place the RF-Plate on a fresh RP-Plate, apply lysates from step 4, and centrifuge for 2 min at 2,500 × g. Exclude steps 6 and 7 and proceed directly to step 8.

6. Load the cell lysates into the wells of the RF-Plate.

7. Apply moderate vacuum (0.2 – 0.4 bar reduction of atmospheric pressure) until all of the samples have passed through the column usually it takes less than 5 min).
   Cover unused wells with a rubber pad if the vacuum does not build up immediately.If using a centrifuge with a swing-bucket rotor capable of hosting 96-well blocks plus the RF-Plate , place the RF-Plate on a fresh RP-Plate, apply lysates from step 4, and centrifuge for 2 min at 2,500 × g.

8. Remove the RP-Plate from the vacuum manifold and add 630µl of isopropanol to each well. Close the block with a RSA-Foil and mix by inverting the block 6 – 8 times. Centrifuge the block for 15 min at ≥ 2,500 × g. If using a centrifuge, spin the block for 10 min at 6,000 × g. Remove foil and discard the supernatant by inverting the block quickly. Tap the block on paper towels.

9. Add 500µl ethanol (70%) per well and seal the block again. Centrifuge at 2,500 × g for 3 min. Remove foil and discard the supernatant again. Tap the block on paper sheet or paper towels.

10. Dry the pellets at 50 – 55°C for 10 – 15 min in an incubator or oven until ethanol is completely removed.

11. Add 50 µl Buffer RE-96 and vortex the block briefly. Alternatively you can use 5mM Tris HCl, pH 8.5.

Optional:
Incubate the block at 50 – 55°C for 10 - 15 min. Centrifuge briefly before removing the foil. For higher DNA concentration the plasmid DNA can be resuspended in 20 µl RE-96.
For downstream applications use approximately ~0.5 µg (for fluorescent dye terminator cycle sequencing) and 1 µg (for fluorescent dye primer)

Automated purification of high-copy plasmid DNA

Pellet the bacteria by centrifuging the RC-Plates (with RG-foil on the block) at 1,000 × g for 10 min (Centrifugation at higher g-forces might produce tight pellets which are more difficult to resuspend)). Pour off the supernatant and tap the block on blotting paper or paper towels in order to remove the remaining medium

Optional:
If centrifugation at higher g-forces is used, a shaker integrated on the robot worktable will be necessary for complete resuspension of the bacterial pellet after addition of Buffer R1-96.

Briefly vortex the RC-Plates on a suitable vortexer before placing in the desired postion of the robot worktableensuring complete resuspension of the pellet after addition of Buffer R1-96.

1. Reconstitute Buffer R1-96 by adding RNase A (see "Preparation of buffers and reagents).

2. Add buffers to the reservoirs or place the buffer bottles in the Corresponding positions of the robot worktable. Place the plastic equipment like plates and the assembled vacuum manifold in the locations as specified in the individual robot program.
   Optional:
   The resuspension buffer RE-96 (5 mM Tris/HCl, pH 8.5) may be substituted by nuclease free water (pH is 8.0–8.5).

3. Select method or program for DNA purification.
   Optional:
   After transfer to the RF-Plate, incubate crude lysates for 1-3 min on the plate. This incubation allows the formation of a compact white precipitate. This step is usually not required for culture volumes up to 1.5 ml.

4. After addition of 0.7 volumes of Isopropanol to each well by the robot remove the RP-Plate from the vacuum manifold.

5. Centrifuge the block for 15 min at ≥ 2,500 × g. Remove foil and discard the supernatant by inverting the block quickly. Tap the block on blotting paper or paper towels. If using a NucleoSwing Z513 centrifuge, spin the block for 10 min at 6,000 × g.

6. Add 500 µl ethanol (70%) per well and seal block again. Centrifuge at 2,500 × g for 3 min. Remove foil and discard the supernatant as described above (step 7). Tap the block on blotting paper or paper towels. Dry the pellets at 50 - 55°C for 10 – 15 min in an incubator or oven until no more ethanol droplets are visible.
   Ethanol is an inhibitor of many enzymatic reactions. Drying of the pellets is also possible at room temperature. However, for optimal removal of ethanol from the preparation, heat incubation is recommended.

7. Add 50 µl RE-96 resuspension buffer (5 mM Tris-HCl, pH 8.5) and vortex the block briefly. Optional: Close the block again with a foil and incubate at 50 – 55°C for 10 - 15 min. Centrifuge briefly before removing the foil. This step is recommended to speed up the resuspension of DNA if you intend to use it for immediately following downstream applications.
   If higher DNA concentrations are desired, the plasmid DNA can be resuspended in 20 µl RE-96 as well. The sealed block can conveniently be used for storage of the plasmid DNA at +4°C or –20°C.

8. Quantify the yield of plasmid DNA by agarose gel electrophoresis before setting up sequencing reactions: compare aliquots of the minipreps to supercoiled plasmid DNA standards with known concentration. We recommend usage of ~1 µg of plasmid DNA for fluorescent dye primer and ~0.5 µg for fluorescent dye terminator cycle sequencing.

Supplementary protocol for automated and manual purification of large low-copy construct DNA

For cultivation of large low-copy constructs, e.g. BACs:

- Use a freshly prepared preculture to inoculate cultures for BAC DNA preparation. Using a preculture gives higher reproducibility and more consistent results.

- Grow pre-cultures in a RP-Plate. Use 150 µl to 1.2 ml of LB or 2 x YT medium with an appropriate antibiotic. Grow with vigorous shaking 300-400 rpm at 37°C for 16 h. Alternatively grow 100 to 150 µl preculture using a standard Microtiter plate (u-bottom) with shaking at 180 rpm. Inoculate precultures with single colonies or glycerol stocks.

Note:
Direct inoculation of cultures for BAC DNA preparation from single colonies or glycerol stocks (without preculture) may result in lower yields and less reproducible results due to higher differences in yields. Use a suitable pin-tool for 96-well plates or 8-channel pipet to inoculate the culture. Avoid repeated freeze/thaw cycles of BAC glycerol stocks. Use replicate plates or prepare fresh glycerol stocks for frequent use.

- Inoculate cultures for BAC DNA peparation from the precultures. Dilute 1 : 1000 (1 µl of preculture per 1 ml of medium with appropriate antibiotic, e.g. 25 µl/ml kanamycin or 12.5 µg/ml chloramphenicol) using a suitable pin-tool or 8-channel pipet.
- When growing BAC cultures in 96-well plates use up to 3 cultures of 1.3 ml per preparation • Alternatively, use suitable 48-well deep well blocks (e.g. ABgene 6 ml storage plate, Whatman Uniplate 48 wells, 5 ml) for culturing BACs. Use 2.5 ml of a suitable culture medium.
- Grow cultures for 16 h at 37°C with shaking. Use the supplied RGP- Foil to cover the culture plate. Discard preculture or prepare new glycerol stocks from preculture.
- Harvest bacteria by centrifugation. Centrifuge culture for 10 min at 2,500 x g. When using more than one culture for preparation of DNA from a clone combine corresponding cultures.
- Use a suitable centrifuge rotor with a clearance of 44 mm to accommodate the square well culture plates. Discard medium after centrifugation by inverting the plate quickly. Remove residual culture medium by tapping the plate on a filter paper.

Note:
Please contact our technical service if you need additional information to adapt the Nucleo-pore Rapid Prep 96 Plasmid kit on common laboratory automation workstations.

1. Centrifuge RC-Plates (with RGP-Foil on the block) at 2,500 × g for 10 min. Remove the Foil and invert the block quickly to pour off the supernatant, ensuring that pellet is not disturbed. Tap the block on a paper sheet or paper towels in order to remove as much of the remaining medium as possible.
2. Add 300 µl buffer R1-96 (RNase added) to each bacterial pellet and resuspend the cells by gentle vortexing till the pellet is dissolved completely.
3. Add 300 µl buffer R2-96 to each well, seal the block thoroughly with RSA-Foil, and mix by gently inverting the block 10 – 15 times. Incubate at room temperature for up to 5 min. Do not vortex. Remove RSA-Foil and dry the upper rim of the wells with a paper towel.
4. Add 300 µl buffer R3-96 to each well, seal the block with fresh RSA-Foil, and mix by gently inverting 10 – 15 times. A white precipitate of SDS and cell debris forms.
5. (Optional Step for endA+ host strains): Seal the block with a fresh RSA-Foil and incubate it in a boiling water bath for 5 min for the heat destruction of endonucleases and denaturationof proteins. Let the block cool down on ice to at least room temperature.

   Note:
   Adjust the vacuum manifold:
   Insert spacers for "RP-Plates", , into the grooves located on the short sides of the manifold with notched side up. Place a fresh RP-Plate into the manifold base, close the lid and insert the RF-Plate into the lid.
6. Load the cell lysates into the wells of the RF-Plate.
7. Apply moderate vacuum (0.2 – 0.4 bar reduction of atmospheric pressure) until all of the samples have passed through the column usually it takes less than 5 min). Cover unused wells with rubber pad.
8. Remove the RP-Plate from the vacuum manifold. Add 630 µl (0.7 volumes) of isopropanol to each well. Close the block with an Adhesive Foil and mix by inverting the block 6 – 8 times.
   Optional: Add 2 mg of glycogen or other carrier for precipitation.
9. Centrifuge the block for 30 min at 6,000 × g at room temperature. (in a refrigerated centrifuge) For centrifugation at lower g-forces centrifugation times can be increased proportionately (e.g. 1hr at 2,000 x g).
10. Remove foil and discard the supernatant by inverting the block quickly. Tap the block on blotting paper or paper towels. Add 500 µl ice-cold ethanol (70%) per well and seal block again. Centrifuge at 6,000 × g for 10 min. For centrifugation at lower g-forces centrifugation times can be increased proportionately (e.g. 1hr at 2,000 x g).
11. Remove foil and discard the supernatant as described in step 10. Tap the block on paper sheet or paper towels. Dry the pellets at room temperature for at least 10 – 15 min until no more ethanol droplets are visible.
12. Add 25-30 µl RE-96 resuspension buffer (5 mM Tris-HCl, pH 8.5), seal the block with RSA-Foil and incubate the block overnight at room temperature.
   Vortexing can be done if DNA is used for sequencing purposes.However, vortexing may cause shearing which might interfere in some mapping applications.

TROUBLE SHOOTING GUIDE

Poor plasmid yield

Possibile Cause
- *No antibiotic (or less amount) used during cultivation*

Suggestion
- Cells harbouring the plasmid of interest may become overgrown by nontransformed cells. Add appropriate amounts of freshly prepared stock solutions to all media, solid and liquid.

Possible Cause
- *Very old cultures*

Suggestion
- Always use fresh cultures as suggested in section Bacterial Culture

Suboptimal performance of plasmid DNA in sequencing reactions, problems with downstream applications

Possible Cause
- *Resuspension of plasmid DNA with TE buffer*

Suggestion
- Repurify the plasmid DNA and elute with nucleasefree water as EDTA contained in Elution BBuffer may inhibit enzymatic reactions like DNA sequencing. Alternatively, the plasmid DNA may be precipitated with ethanol, and redissolved in RE-96 buffer or nuclease-free water.

Possible Cause
- *Quantity of DNA used for sequencing reactions is low*

Suggestion
- Quantitate DNA by agarose gel electrophoresis before setting up sequencing reactions.

Possible Cause
- Contamination of final plasmid preparation with ethanol

Suggestion
- Insufficient drying after washing with ethanol, and therefore remaining ethanol may cause problems with downstream applications like DNA sequencing or loading of samples onto agarose gels.

High level contamination with chromosomal DNA

Possible Cause

- *Excessive mixing steps and vortexing after addition of lysis buffers buffer R2-96 and buffer R3-96, or before transfer of lysate to the RF- Plate.*

Suggestion

- *Always mix by gentle inversion of the sealed culture block.*

Possible Cause

- *Culture volume was too high*

Suggestion

- Reduce culture volume if lysate is too viscous for gentle and complete mixing.

Possible Cause

- *Bacterial culture overgrown*

Suggestion

- Overgrown bacterial cultures contain lysed cells and degraded DNA. See suggestions in section 5 'Growing of bacteria cultures'.

Possible Cause

- *Lysis was too long*

Suggestion

- Lysis step must not exceed 5 min.

Incomplete lysis of bacterial cells

Possible Cause

- *Cell pellet not properly resuspended*

Suggestion

- It is essential that the cell pellet is completely resuspended prior to lysis. No cell clumps should be visible before addition of lysis Buffer R2-96. If necessary, increase number of mixing cycles or duration of shaking.

Possible Cause

- *SDS in Buffer R2-96 precipitated*

Suggestion

- SDS in Buffer R2-96 may precipitate upon storage. If this happens, incubate BUFFER R2-96 at 30–40°C for 5 min and mix well.

Possible Cause

- *Too many bacterial cells used*

Suggestion

- Usage of LB as the growth medium is recommended. When using rich media like TB or 2 x YT, cultures volumes have to be reduced

ORDERING INFORMATION

Description	Pack Size	Cat. No.
® DNASure Tissue Mini Kit	50 preps	NP-61305
® DNASure Plant Mini Kit	50 preps	NP-79105
® DNASure Plant Mini Kit	250 preps	NP-79107
® DNASure Plant Midi Kit	20 preps	NP-78153
® DNASure Plant Maxi Kit	10 preps	NP-78164
® DNASure Blood Mini Kit	50 preps	NP-61105
® DNASure Blood Mini Kit	250 preps	NP-61107
® DNASure Blood Midi Kit	20 preps	NP-61184
® DNASure Blood Maxi Kit	10 preps	NP-61193
® DNASure Blood FastPure Kit	50 preps	NP-62205
® DNASure Blood FastPure Kit	250 preps	NP-62207
® SureSpin Plasmid Mini Kit	50 preps	NP-37105
® SureSpin Plasmid Mini Kit	250 preps	NP-37107
® SureSpin Plasmid FastPrep Kit	50 preps	NP-47105
® SureSpin Plasmid FastPrep Kit	250 preps	NP-47107
® SureSpin Buffer Set*	1	37107-BS
® SurePrep Plasmid Mini Kit	20 preps	NP-15123
® SurePrep Plasmid Mini Kit	100 preps	NP-15125
® SurePrep Plasmid Midi Kit	20 preps	NP-15143
® SurePrep Plasmid Midi Kit	100 preps	NP-15145
® SurePrep Plasmid Maxi Kit	10 preps	NP-15161
® SurePrep Plasmid Maxi Kit	25 preps	NP-15162
® SurePrep Plasmid Mega Kit	5 preps	NP-15183
® SurePrep Plasmid Giga Kit	5 preps	NP-15191

***SureSpin® Buffer Set**

For the isolation of low-copy plasmids, buffers PA1, PA2, PA3, RNase A, sufficient for 300 preps

ORDERING INFORMATION

Description	Pack Size	Cat. No.
SurePrep® Buffer Set**	1	15143-BS
SurePrep® Plasmid Endofree Maxi Kit	10 preps	NP-15363
SurePrep Plasmid Endofree Mega Kit	5 preps	NP-15365
SurePrep® Plasmid Endofree Giga Kit	5 preps	NP-15367
SureSpin® 96 PCR Kit	4x96	NP-38151
SureTrap® Gel Extraction Kit	50 preps	NP-38705
SureTrap® Gel Extraction Kit	250 preps	NP-38707
SureTrap® PCR Cleanup Kit	50 preps	NP-38105
SureTrap® PCR Cleanup Kit	250 preps	NP-38107
SureExtract® Spin PCR/Gel Extraction Kit	50 preps	NP-36105
SureExtract® Spin PCR/Gel Extraction Kit	250 preps	NP-36107
SureSEQ® Cleanup Kit	50 preps	NP-73205
RNASure® Mini Kit	50 preps	NP-84105
RNASure® Mini Kit	250 preps	NP-84107
RNASure® Plant Kit	50 preps	NP-84905
RNASure® Plant Kit	250 preps	NP-84907
miRNASure® Mini Kit	50 preps	NP-71002
SureTrap® mRNA Mini Kit	12 preps	NP-80033
SureTrap® mRNA Midi Kit	12 preps	NP-80043
RNASure® Virus Kit	50 preps	NP-67705
RNASure® Virus Kit	250 preps	NP-67707

****SureSpin® Buffer Set**

For isolation of low-copy plasmids, cosmids, BACs, PACs, and P1 constructs, only applicable with SurePrep® Plasmid kits, sufficient for 10 SurePrep Maxi Columns (Maxi preps), 20 SurePrep® Midi Columns (Midi preps), set incl. RNase A

ORDERING INFORMATION

Description	Pack Size	Cat. No.
Nucleo-pore® Stool DNA Mini Kit	50	NP-7011D
Nucleo-pore® gRNA Blood Kit	50	NP-0201R
Nucleo-pore® gDNA Urine Kit	20	NP-6030D
Nucleo-pore® Yeast Transformation Kit	120	NP-1002T
Nucleo-pore® DNA Methylation Kit	50	NP-6006D
Nucleo-pore® gDNA Clean-up Kit	200	NP-4304D
Nucleo-pore® Bisulphite DNA Clean-up Kit	50	NP-5205D
Nucleo-pore® gDNA Fungal/Bacterial Mini Kit	50	NP-7006D

Product Warranty

Nucleo-pore™ RapidPrep 96 plasmid kit components are intended for research purposes only. They are suitable for in - vitro uses only. The purchaser must determine the suitability of the product for its particular use. Should any product fail to perform satisfactorily due to any reason other than misuse, Genetix will replace it free of charge or refund the purchase price. Genetix reserve the right to change, alter, or modify any product to enhance its performance and design. It is the responsibility of the user to verify the use of the Nucleo-pore RapidPrep 96® plasmid kit for a specific application range as the performance characteristic of this kit has not been verified to a specific organism. No claim or representation is intended for its use to identify any specific organism or for clinical or therapeutic use.

Genetix does not warrant against damages or defects arising in shipping and handling (transport insurance for customers excluded), or out of accident or improper or abnormal use of this product.

In accordance with Genetix ISO-certified Quality Management System, each lot of Nucleo-pore RapidPrep 96® plasmid kit is tested against predetermined specifications to ensure consistent product quality.

In no event shall Genetix be liable for claims for any other damages, whether direct, indirect, incidental, compensatory, foreseeable, consequential, or special (including but not limited to loss of use, revenue or profit), whether based upon warranty, contract, tort (including negligence) or strict liability arising in connection with the sale or the failure of Genetix products to perform in accordance with the stated specifications.

Product claims are subject to change. Therefore please contact our Technical Support Department for updated information on Genetix products.

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